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|---|---|--|
| <b>CID</b> : 2110644187                             | <b>SID</b> : 177803369199               | <b>R<br/>E<br/>P<br/>O<br/>R<br/>T</b> |
| <b>Name</b> : Yashmeet Rout                         | <b>Registered</b> : 19-Sep-2021 / 16:54 |  |
| <b>Age / Gender</b> : 7 Years / Male                | <b>Collected</b> : 19-Sep-2021 / 17:11  |  |
| <b>Dr.</b> :  | <b>Reported</b> : 20-Sep-2021 / 15:35   |  |
| <b>Reg. Location</b> : Kandivali East (Main Centre) | <b>Printed</b> : 20-Sep-2021 / 15:55    |  |

**Real time Qualitative RT-PCR detection of 2019-nCOV RNA / COVID-19 RNA**

| <b><u>PARAMETER</u></b>  | <b><u>RESULT</u></b>                |
|--|-------------------------------------|
| Result   | SARS-CoV-2: Not Detected (Negative) |
| Kit: Viral Detect II Multiplex(Genes2Me), Target gene(RdRP & N), Cutoff: <37 |                                     |

**ICMR Registration No: Andheri-Mumbai- SUBUR001, Pune-SUDIPLPMH**

**Specimen: Nasopharyngeal & Oropharyngeal swab in VTM Method: Real time RT-PCR**

**Note:**

- Ct value indicates the infectivity and not severity of infection.
- ICMR recommended kits are used for reporting. All the positive cases will be notified to ICMR for further surveillance.
- Clinical correlation with patient history, radiology findings and co-infection with other virus infection is necessary to be determined especially in cases with Borderline positive Ct values.
- Borderline positive cases (Ct Value >30) may give variable results on repeat testing. The possible reasons could be the variations in kits and instruments used.

**Limitations:**

- Optimum specimen types and timing of peak viral levels during infections caused by 2019-nCOV have not been determined. Collection of multiple specimens (Types & Time points) may be necessary in view of suspected clinical history. The repeat specimen may be considered after a gap of 2-4 days after the collection of first specimen for additional testing if required. (other respiratory pathogens)
- Negative results do not preclude SARS - CoV - 2 infection and should not be used as the sole basis for patient management decisions.
- This test is a qualitative assay and does not quantify viral load. Various host factors, viral factors, variability in sample collection / site and techniques used by the laboratories can affect Ct values. Therefore, Ct values are not an absolute indication of viral load and should be interpreted with caution.

**Factors leading to false negative RT-PCR report:**

- Inadequate specimen collection, Poor quality of sample and non-representative sample.
- Sample collected too early or too late in the infection, Improper sample handling and shipment.
- Technical reasons- PCR Inhibitor, analytical sensitivity of kit used.
- Active recombination &/ mutations in target genes used for detection of SARS-CoV-2 virus.

**References:**

1. Diagnostic detection of 2019-nCoV by real-time RT-PCR, Berlin Jan 17th, 2020.
  2. Labcorp COVID-19 RT-PCR test EUA Summary / COVID-19 RT-PCR test (laboratory corporation of America).
- \* Sample processed at Molecular Diagnostics Laboratory, CPL, Andheri West  
\*\*\* End Of Report \*\*\*

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