



<b>CID</b> : 2110644133	<b>SID</b> : 177803369166	<b>R E P O R T</b>
<b>Name</b> : MR.DIPAK KUMAR ROUT	<b>Registered</b> : 22-Jan-2021 / 09:44	
<b>Age / Gender</b> : 41 Years / Male	<b>Collected</b> : 22-Jan-2021 / 09:54	
<b>Dr.</b> : -	<b>Reported</b> : 22-Jan-2021 / 17:30	
<b>Reg. Location</b> : Kandivali East (Main Centre)	<b>Printed</b> : 22-Jan-2021 / 20:06	

**Real time Qualitative RT-PCR detection of 2019-nCOV RNA / COVID-19 RNA**

<b>PARAMETER</b>	<b>RESULT</b>
Result	SARS-CoV-2: Not Detected (Negative)

Kit:Viral Detect II Multiplex(Genes2Me),Target gene(RdRP & N),Cutoff:<37

ICMR Registration No: Andheri-Mumbai- SUBUR001, Pune-SUDIPLPMH

Specimen: Nasopharyngeal & Oropharyngeal swab in VTM Method: Real time RT-PCR

**Note:**

- Ct value indicates the infectivity and not severity of infection.
- ICMR recommended kits are used for reporting. All the positive cases will be notified to ICMR for further surveillance.
- Clinical correlation with patient history, radiology findings and co-infection with other virus infection is necessary to be determined especially in cases with Borderline positive Ct values.
- Borderline positive cases (Ct Value >30) may give variable results on repeat testing. The possible reasons could be the variations in kits and instruments used.

**Limitations:**

- Optimum specimen types and timing of peak viral levels during infections caused by 2019-nCOV have not been determined. Collection of multiple specimens (Types & Time points) may be necessary in view of suspected clinical history. The repeat specimen may be considered after a gap of 2-4 days after the collection of first specimen for additional testing if required. (other respiratory pathogens)
- Negative results do not preclude SARS - CoV - 2 infection and should not be used as the sole basis for patient management decisions.
- This test is a qualitative assay and does not quantify viral load. Various host factors, viral factors, variability in sample collection / site and techniques used by the laboratories can affect Ct values. Therefore, Ct values are not an absolute indication of viral load and should be interpreted with caution.

**Factors leading to false negative RT-PCR report:**

- Inadequate specimen collection, Poor quality of sample and non-representative sample.
- Sample collected too early or too late in the infection, Improper sample handling and shipment.
- Technical reasons- PCR Inhibitor, analytical sensitivity of kit used.
- Active recombination &/ mutations in target genes used for detection of SARS-CoV-2 virus.

**References:**

1. Diagnostic detection of 2019-nCoV by real-time RT-PCR, Berlin Jan 17th, 2020.
2. Labcorp COVID-19 RT-PCR test EUA Summary / COVID-19 RT-PCR test (laboratory corporation of America).

\* Sample processed at Molecular Diagnostics Laboratory, CPL, Andheri West  
\*\*\* End Of Report \*\*\*

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